Spongiforma, a new genus of gasteroid boletes from Thailand

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Based on morphological and molecular characters, *Spongiforma* is described as a new genus of gasteroid boletes belonging in the Boletineae. It is represented by a single species, *S. thailandica*, that is putatively mycorrhizal with dipterocarp trees in central Thailand. Unusual morphological features include a sponge-like, astipitate, epigeous basidiome with large exposed locules and a strong coal tar odor, and rugulose, reddish brown basidiospores with an apical pore that become smooth and violet grey in 3% potasium hydroxide solution. A description, illustrations, phylogenetic analysis and comparison with allied taxa are presented.

Key words: Agaricomycotina, Basidiomycota, Boletineae, molecular phylogenetics, taxonomy.

Article Information

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Introduction

Although Southeast Asia is home to numerous genera of boletes (Basidiomycota, Agaricomycotina, Boletales), few secotioid or gasteroid taxa have been reported from the region (Corner and Hawker, 1953; Corner, 1972; Watling and Lee, 1995, 1998; Lee et al., 2002; Halling et al., 2007). Recently, specimens of an unusual, epigeous, astipitate and sponge-like species were collected from Khao Yai National Park in central Thailand. Basidiomes have a rubbery-gelatinous texture, emit a strong coal tar odor, and are associated with Shorea henryana Pierre and Dipterocarpus gracilis Blume (Dipterocarpaceae) in a primary forest. A unique set of macro- and micromorphological features in combination with molecular sequences of the nuclear large subunit (nuc-lsu) gene region indicate that the organism represents a new genus in the Boletineae. A description, illustrations, phylogenetic analysis and comparison with allied taxa are presented herein.

Materials and Methods

Morphology

Macromorphological data were derived from fresh specimens, whereas micromorphological data were derived from dried specimens rehydrated in ethanol followed by distilled water, 3% KOH or Melzer's reagent. Duplicate specimens are deposited in BBH and SFSU.

DNA extraction, PCR and DNA sequencing

Collections used in this study for molecular analyses are listed in Table 1. Genomic DNA from herbarium specimens was isolated following the phenol/chloroform procedure by Lee and Taylor (1990). The crude DNA extracts were diluted up to 1000-fold with deionized water for use as PCR templates. PCR reactions were performed for two nuclear rDNA regions using the primer combinations ITS1-F-ITS4 (ITS region including the 5.8S gene) and LR0R-LR5 (nuc-lsu). Sequences of primers used in this study have been described elsewhere (Vilgalys and Hester, 1990; White

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Table 1. Isolate numbers, location, date, collector and accession numbers for fungal isolates sequenced for use in this study.

Species	Isolate	Origin	Date	leg./det.	GenBank accession numbers	
					ITS	nuc-lsu
Chalciporus rubinellus	191/81	U.S.A., ME	8/1981	W. Steglich	EU685111	EU685106
Porphyrellus pseudoscaber	RH8716	U.S.A., CA	11/15/2005	R. Halling	EU685112	EU685107
Spongiforma thailandica	DED7873	Thailand	7/7/2005	D. Desjardin	EU685113	EU685108
Strobilomyces sp.	RH4514	Australia	2/1992	R. Halling		EU685109
Tylopilus atronicotianus	Both s.n.	U.S.A., NY	9/18/2004	E. Both	EU685114	EU685110

et al., 1990; Gardes and Bruns, 1993). The amplifications were run in 35 cycles on a PTC-200 thermal cycler (MJ Research, Waltham, Massachusetts) using the following parameters: denaturation 94°C (1 min), annealing 50°C (45 sec), extension 72°C (1.5 min). PCR products were purified using Pellet Paint (Novagen, EMB Biosciences, San Diego, California). All PCR products were sequenced using BigDye terminator sequencing 3.1 chemistry (Applied Biosystems, Foster City, California), and run on an Applied Biosystems 3130 genetic analyzer. Contiguous sequences were assembled and edited using Sequencher 4.7 (GeneCodes Corp., Ann Arbor, Michigan). Automated alignments that were generated with ClustalX (Thompson et al., 1997) were manually adjusted in MacClade 4.08 (Maddison and Maddison, 2005).

Datasets and phylogenetic analyses

Initial blast searches (Altschul et al., 1997) using the ITS sequence of S. thailandica as a query produced exclusively hits in the Boletaceae, however, without providing significant results at the generic rank. The newly generated nuc-lsu rDNA data were first blasted and then, along with the best blast hits, aligned into the dataset of Binder and Hibbett (2006), which was narrowed down to 85 taxa after several rounds of consecutive bootstrapping and pruning of redundant sequences. The final alignment included 933 positions and the data were analyzed using maximum-parsimony (MP), maximum-likelihood (ML) and Bayesian methods. A non-parametric MP bootstrap analysis was performed using 1000 replicates PAUP*4.0b10 (Swofford, 2002). characters equally weighted, one random taxon addition sequence, and tree bisection reconnection (TBR) branch swapping. RAxML v. 2.2.3 (Stamatakis, 2006) was used to run 100 ML replicates under the GTRMIX model and, in addition, 1000 ML bootstrap replicates under the GTRCAT model. A 50% majorityrule consensus tree was built from the resulting 1000 trees in PAUP* to estimate confidence values. Finally, posterior probabilities (PP) were determined in two independent analyses by running one cold and three incrementally heated Metropolis-coupled Markov chain Monte Carlo (MCMCMC) analyses for 3×10^6 generations using MrBayes v3.0b4 (Ronquist and Huelsenbeck, 2003), saving trees every 100^{th} generation. The GTR+ Γ +I model was specified as prior, assuming equal probability for all trees and unconstrained branch length. The final burn-in period determining the stationary proportion of trees saved after likelihood scores converged to a stable equilibrium was estimated using Tracer v1.4 (Rambaut and Drummond, 2007). A 50% majority-rule consensus tree was generated from the stationary trees in PAUP* to estimate confidence intervals. The results of the analyses are summarized in Fig. 1.

Results

Phylogenetic analyses

The complete ITS sequence obtained from *Spongiforma thailandica*, consisting of the two spacer regions and 5.8S gene, has a size of 682 bp. Blast searches using the entire ITS sequence and the three regions separately as queries produce only matches for the 5.8S gene, and place the species in the Boletaceae.

Inferences on the nuc-lsu dataset under parsimony, likelihood, and Bayesian methods yielded highly similar tree topologies,

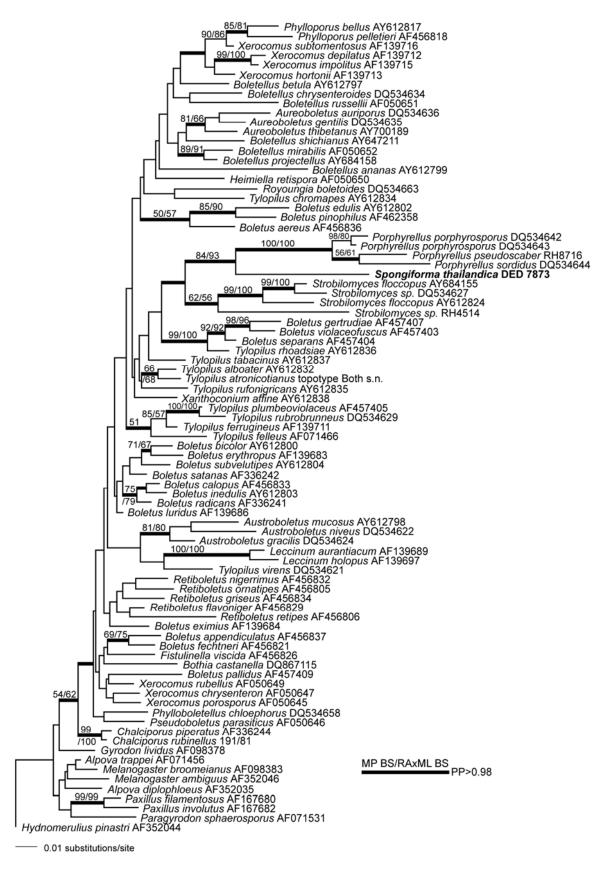


Fig. 1. Phylogenetic relationships of *Spongiforma* inferred from nuc-lsu rDNA data using RAxML. One of 100 trees is shown here ($-\ln L = 10299.869$) and BS support values > 50% are indicated along nodes, while PP > 0.98 are marked as thickened branches. A GenBank accession number is provided for published sequences and the isolate number is provided for newly generated data.

suggesting that the new genus Spongiforma is the sister group of Porphyrellus. Long branches produced by species in both genera seem to obscure this relationship to some degree. Nevertheless, high statistical support received from three different methods suggests that this finding is based on a true phylogenetic signal and not a result of long-branch attraction. In addition, the independent Bayesian runs under a $GTR+\Gamma+I$ model, which predetermined (Binder & Hibbett 2006) without testing the suitability of simpler models for this analysis, repeatedly and quickly converged after 230 000 generations. Thus, the clade including Spongiforma and Porphyrellus consistently resolved in all analyses presented here (Fig. 1).

Taxonomy

Spongiforma Desjardin, Manf. Binder, Roekring & Flegel gen. nov.

MycoBank: MB 512541

Etymology: Referring to the sponge-like basidiomes.

Type species: Spongiforma thailandica Desjardin, Manf. Binder, Roekring & Flegel

Basidiomata epigaea, sessilia, cerebriformia. Peridium nulla. Gleba loculis labyrinthiformibus, 2-20 mm lata, brunneis vel rubrobrunneis; columella irregularis, pyriformis, cremea-alba. Basidiosporae amygdaliformae, symmetricae, rugulosae, apiculatae cum poro, inamyloideae, cyanophileae. Basidia statismosporicae, 4-sporigera. Cystidia cylindrica vel ventricosa-rostrata. Trama glebae gelatinosae. Fibulae nulla.

Basidiomes epigeous, sessile, cerebriform to sponge-like with rubbery-gelatinous texture; peridium absent; locules 2-20 mm diam., irregular in outline, lined with a smooth, grevish orange to brown or reddish brown hymenium, sterile ridges white to creamcoloured; columella poorly developed, pyriform, cream-coloured, attached to white rhizomorphs. Basidiospores brown to vinaceous brown in mass, amygdaliform, bilaterally symmetrical, rugulose, with a central apiculus and small apical pore, reddish brown in water, violet grey in 3% KOH, inamyloid, cyanophilic. Basidia statismosporic, 4-sterigmate. Cystidia common on sterile locule edges, cylindrical to ventricose-rostrate, hyaline. Tramal hyphae gelatinous, inamyloid. Clamp connections absent.

Spongiforma thailandica Desjardin, Manf. Binder, Roekring & Flegel **sp. nov.**

(Fig. 2a-f)

MycoBank: MB 512542

Etymology: Referring to the country in which the species grows.

Basidiomata epigaea, sessilia, cerebriformia, 50-100 × 40-70 mm, globosa vel ovoidea. Peridium nulla. Gleba loculis labyrinthiformibus, 2-20 mm lata, brunneis vel rubrobrunneis; columella irregularis, pyriformis, cremea-alba. Basidiosporae (9–)10–11.5(–12.5) × 5.5-7(–7.5) μm, amygdaliformae, symmetricae, rugulosae, apiculatae cum poro, inamyloideae, cyanophileae. Basidia statismosporicae, 25–32 × 6.5–9.5 μm, cylindrica vel subclavata, 4-sporigera, sterigmata usque 9.5 μm. Cystidia 25–48 × 5–10 μm, cylindrica vel ventricosa-rostrata. Trama glebae gelatinosae. Fibulae nulla. Holotypus hic designatus: Thailand, DED 7873 (BBH)

Basidiomes (Fig. 2a-b) epigeous, 50–100 mm diam \times 40–70 mm tall, astipitate, irregularly globose to ovoid, cerebriform to sponge-like, rubbery-pliant (not brittle; can be squeezed like a sponge to remove water and it will spring back to its original shape). *Peridium* absent. Hymenophore composed of ridges or folds delimiting empty locules; locules 2–15(– 20) mm diam, irregular in outline, lined with a well-developed hymenium, pale greyish orange (5B3-4) to brownish grey (6C3) when young, becoming light brown (6D4-5) to brown (6E5-8), dark brown (6F5-8) or reddish brown (8-9E6-8) at maturity; sterile ridges paler, white to cream when dried; all tissues immediately deep purple to purplish black in 10% KOH. Spores brown (6E5-6) to light brown (6D5-6) or vinaceous brown (8E-F6). Columella 10-15 mm tall \times 8–10 mm diam (apex) \times 3–4 mm diam (base), obconical to pyriform, apex lacunose, base folded, cream buff (4A3) to pale grevish orange (5B3); attached to fine, reticulate-branched, white rhizomorphs. Odor mild and pleasant when young, but soon becoming strong and unpleasant, of coal tar or burned rubber (like Tricholoma sulphureum), can be detected from 10 m distant. Peridium absent. Sterile ridges composed of erect chains of cells terminated by cystidia. Cystidia 25–48 × 5-10 µm, cylindrical to subclavate or ventricose-rostrate, obtuse, hyaline or a few with pale brown cellular contents, inamyloid, thin-walled; abundant on the sterile locule edges and scattered amongst basidia near the

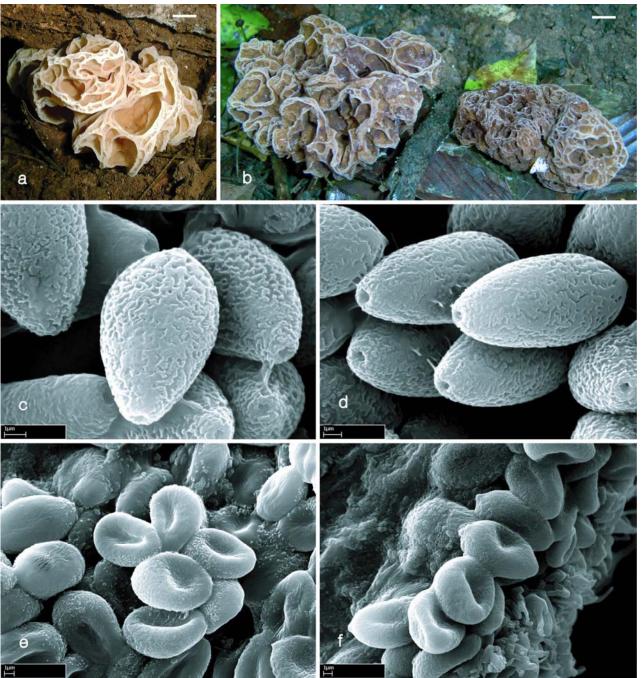


Fig. 2. Spongiforma thailandica. a-b. Basidiomes (DED 7485). c-d. SEM of air dried basidiospores (DED 7873, holotype). Note the distinct ornamentation and obvious apical pore. e-f. SEM of basidiospores mounted in 3% KOH (DED 7873, holotype). Note the loss of an apical pore and nearly smooth surface. — Scale bars: a-b = 10 mm; c-f = 1 μ m.

locule edges but absent elsewhere. *Hymenophoral trama* of subparallel to slightly interwoven hyphae 3–10 μ m diam, cylindrical, branched, septa often inflated (like an elbow joint), gelatinous, hyaline, inamyloid, thinwalled. *Subhymenium* of inflated to vesiculose cells 9–20 × 9–14 μ m, hyaline, inamyloid, thinwalled, non-gelatinous. *Hymenial cystidia* absent (except as noted above). *Basidia* statismosporic, 25–32 × 6.5–9.5 μ m, cylindrical to subclavate, 4-spored with straight

sterigmata up to 9.5 µm long, unclamped. Basidioles cylindrical to subclavate. Basidiospores (Fig. 2c-f) (9–)10–11.5(–12.5) \times 5.5-7(–7.5) µm [xmr = 10.7–11.1 \times 5.9–6.5 µm, xmm = 11 \pm 0.3 \times 6.2 \pm 0.4, Q = 1.5–2.1, Qmr = 1.7–1.8, Qmm = 1.76 \pm 0.09, n = 20-25 spores per 2 specimens], amygdaliform, broadest at proximal end and gradually narrowed towards distal end, bilaterally symmetrical, with a small central apiculus, subtruncate at distal end and forming a tiny, narrow pore that may have a

small lip, walls thick ($-0.5 \mu m$), thinner at the distal end, roughened to rugulose overall and reddish brown in water (Fig. 2c-d), appearing nearly smooth and violet grey in 3% KOH (Fig. 2e-f), inamyloid, cyanophilic. *Clamp connections* absent in all tissues.

Habit and habitat: Solitary, in soil under Dipterocarpus sp. and Shorea sp. in primary forest.

Known distribution: Thailand.

Material examined: THAILAND, Nakorn Nayok Province, Khao Yai National Park, Princess Trail ca 2 km from Visitor Center, N14°26.142', E101°23.080', elev. ca 750 m, 9 July 2002, *E. Horak, T. Flegel and D.E. Desjardin* as DED 7485 (BBH, SFSU); same location, 7 July 2005, *D.E. Desjardin*, DED 7873 (holotype BBH, isotype SFSU).

Notes: Diagnostic features of Spongi-forma thailandica include the following: a relatively large, sponge-like and rubbery basidiome colored pale brownish grey to brown or reddish brown with irregular, relatively large locules lined with sporogenous tissue; no stipe but a small columella that is attached to copious, fine white rhizomorphs; a strong coal tar odor (like Tricholoma sulphureum); tissue that turns purple in 3-10% KOH; amygdaliform basidiospores with a tiny apical pore that are reddish brown and rugulose-roughened in water (Fig. 2c-d) but become violet grey, collapsed and nearly smooth in KOH (Fig. 2e-f); and the absence of clamp connections.

Discussion

The new Thai taxon is superficially similar to Gymnopaxillus nudus Claridge, Trappe & Castellano, a species described from material associated with *Eucalyptus* Australia (Claridge et al., 2001). In common with Spongiforma, Gymnopaxillus nudus forms astipitate, cerebriform, bright ferrugineous basidiomes that lack a peridium and have relatively large locules, a basal columella, white rhizomorphs and cyanophilic, bilaterally symmetric basidiospores. However, the genus Gymnopaxillus forms smooth basidiospores that are ellipsoid to subfusoid, golden yellow in KOH and lack a tiny apical pore. In addition, G. nudus differs in forming primarily hypogeous basidiomes, forms a larger and more welldeveloped columella, lacks a strong odor, lacks purple KOH reaction, has longer

basidiospores (11-16 µm), and is mycorrhizal with *Eucalyptus*. Molecular data indicate that *G. nudus* is allied with *Austropaxillus* species (Jarosch 2001) and together they are sister of *Serpula* in the Serpulaceae, all distantly related to *Spongiforma* (Binder and Hibbett, unpubl. data).

Our results drawn from the nuc-lsu dataset indicate that Spongiforma belongs in Boletaceae where it is Porphyrellus (with 84% MP BS and 93% RAXML BS; PP = 1.0), although on a long branch. Together, this group forms the sister clade of Strobilomyces (Fig. 1), which is resolved in all analyses but is only supported by PP = 0.98. As noted in previous studies, ITS sequences of Porphyrellus and Strobilomyces spp. are highly divergent (Lutzoni et al., 2004), and we were not able to align both taxa to the S. thailandica sequence or the latter sequence to any other taxa in the Boletaceae.

Bolete species with reddish brown to vinaceous brown, rugulose to perforatepunctate basidiospores have been placed historically in a number of taxa including Boletus subgen. Tylopilus (Corner, 1972), Tylopilus subgen. Porphyrellus, Austroboletus (both Wolfe, 1979), or Porphyrellus (Singer, 1945; Wolfe and Petersen, 1978). Two Southeast Asian species with basidiospore features very similar to those of Spongiforma thailandica are currently named Austroboletus tristis (Pat. & C.F. Baker) Wolfe and A. longipes (Massee) Wolfe. All three latter species form basidiospores with rugulose to rugulose-punctate surface that turn purple to violaceous grey in KOH. SEM photos of the type specimens of A. tristis and A. longipes (Figs 1, 2 and 4, respectively in Wolfe and Petersen, 1978) clearly show ornamentation similar to that of S. thailandica (Fig. 2c-d) although an apical pore was not reported on basidiospores of any Austroboletus species. Recently collected material of A. longipes from Malaysia confirms these observations (Chan & Halling, NYBG, pers. comm.). Sequences of A. tristis and A. longipes were not available for inclusion in our analyses. It is interesting to note, however, that in our analyses (Fig. 1), Spongiforma was more closely allied with the smooth-spored Porphyrellus (P. porphyrosporus (Fr.) E.-J. Gilbert, P. pseudoscaber Secr. ex Singer, P. sordidus (Frost) Snell) than with the rugulose-punctate-spored Austroboletus (A. mucosus (Corner) Wolfe, A. niveus (G. Stev.) Wolfe, A. gracilis (Peck) Wolfe). An apical pore at the distal end of basidiospores is a rare feature in the boletes. Thiers (1975) reported the basidiospores of *Porphyrellus amylosporus* A.H. Sm. as dark reddish brown, smooth and truncate with a thin-walled depression. Several World species of Heimioporus (= Heimiella) form basidiospores with apical pores, such as Heimiella sp. 2 as illustrated in two SEM photographs by Watling and Hollands (Figs 11-12; 1989), and H. fruticicola (Berk.) E. Horak (R. Halling, NYBG, pers. comm.). In our molecular analyses (Fig. 1), Heimioporus is distantly related to Spongiforma. Another interesting similarity is that the pileipelli of Austroboletus and Porphyrellus species is a (ixo-) trichodermium formed from chains of short, cylindrical to submoniliform cells with cylindrical to clavate terminal cells. This anatomy is similar to that of the sterile edges of the tissues that delimit the locules in Spongiforma, which may represent vestigial pileipellis (peridium) tissue.

Spongiforma thailandica was first reported from Thailand as *Hymenogaster* sp. and illustrated with two color photographs in Ruksawong and Flegel (2001: 248).

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