Sporal characters in *Gomphales* and their significance for phylogenetics

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Traditionally, sporal characters, such as color, shape and ornamentation, have been important in differentiating the various genera within the *Gomphales*. In some instances, however, no precise analyses have been made that would allow us to build primary homologies between these and other spore features. For this study, the characteristics of the basidiospores of 14 taxa of *Gomphales* were examined, using both photonic and electronic microscopy. These examinations clearly demonstrated that spore ornamentation is a very variable character and data, such as the base shape of the spore and the hilar appendix, previously not considered in the taxonomy of this group, can be very informative at this level.

Key words: basidiospores, *Gomphales*, spore ornamentation.

Introduction

The selection of homologous characters is a fundamental process within any systematic study, although there is a great debate regarding their recognition at different levels of organization (Patterson, 1982; Roth, 1988; Hall, 1994; Brower, 2000; Hawkins, 2000). Phylogenetic homology is the historical link between structures or processes of different organisms that share a common ancestry and is the most relevant concept in systematics and comparative anatomy. Since homology allows us to recognize historical connections between modified structures in organisms, phylogenetic homology is pivotal in the study of patterns of similarity at various levels. Taxonomy based on the formal study of characters as a hypothesis of a homology is the best framework for understanding biological diversity (De Luna and Mishler, 1996).

Homology cannot be assessed directly, but needs to be inferred through several analytical approaches that permit the study of the variation patterns of

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discernible features from different viewpoints and using various techniques. Cladistic methods, for example, provide criteria and explicit procedures to determine homologies (Roth, 1988; Bryant, 1989; Mabee, 1989; De Pinna, 1991; Hall, 1994; De Luna and Mishler, 1996; Rieppel and Kearney, 2002).

Spores, like any other fungal structures, contain potential phylogenetic information, and the comparison of characters allows one to arrive at an initial conjecture of homology (primary homology). Despite the fact that systematists have shown the greatest interest in spores when proposing relationships between fungal groups at different taxonomic levels (Pegler and Young, 1971, 1979, 1981; Kühner, 1976), only a few sporal characters have been considered in the descriptions of families comprising the traditional order *Aphyllophorales* (Donk, 1964). Nevertheless, a few of these characters have been used for quite a long time.

Donk (1961, 1964) described the Gomphaceae for the genera Beenakia, Gomphus, Kavinia, Lentaria and Ramaria. One of the characters that allowed him to differentiate this group from others within the Apyllophorales was the presence of almost invariably ellipsoid, oblong or amygdaliform spores, often ochre-colored and only rarely colorless, not amyloid and with variable ornamentation (rarely smooth), which vigorously absorbed cotton blue. Jülich (1981) proposed that the Gomphales order consisted of four families Gomphaceae, Lentariaceae and (Beenakiaceae, Ramariaceae). emphasizing the presence of yellow-ochre, ellipsoid spores. The majority of these spores displays a cyanophilic ornamentation and are rarely smooth. The spore characters of the *Lentariaceae* are particularly different from those of the remaining Gomphales members since they vary from being elongated, ellipsoid to cylindrical or sigmoid and are generally hyaline. The spore wall is thin and slightly amyloid in a few taxa, but never dextrinoid or cyanophilic. Nevertheless, other features, such as the hyphal construction of the basidia, suggest a close affinity with Ramariaceae, similarities that had been already questioned by Corner (1950, 1970). These findings, in turn, led some authors, such as Petersen (1988) and Estrada-Torres (1994), to reconsider Gomphaceae sensu Donk (1961) and to propose the exclusion of *Lentaria* from this group.

Maas Geesteranus (1963) reported that the spores of *Beenakia dacostae* (*Beenakiaceae*) are very similar to those of *Ramaria* (*Ramariaceae*) in terms of their color, ornamentation and form of the hilar appendix. Similarly, Jülich and Star (1983), after considering the ultrastructure of the basidiospores of *B. dacostae*, concluded that it was very similar to others in the *Gomphales*, such as *Gomphus* and *Ramaria*.

Petersen (1967, 1971a, 1981) and Petersen and Pearman (1973) demonstrated the existence of a wide margin of spore ornamentation within

Ramaria and *Gomphus*, and discussed the similarity of the spores between these two genera.

The order Gomphales was also recognized by Hawksworth et al. (1995), with modifications for the Ramariaceae (including also Delentaria, Kavinia and Ramaricium), leaving Lentaria as the sole member of the Lentariaceae family. Hawksworth et al. (1995) included Beenakia as the only member of the Beenakiaceae, and transferred it to the Boletales. He may have arrived at this idea because of the similarities between Beenakia and some Boletales in terms of the color and shape of their spores, similarities that are also found between Austroboletus, Strobilomyces and several members of Gomphaceae sensu Donk (Petersen, 1971a; Pegler and Young, 1981). The genera Pseudogomphus and Terenodon (described later) are also included in the Gomphaceae (Heim, 1970; Maas Geesteranus, 1971). Recent systematic investigations (Hibbett et al., 1997; Pine et al., 1999; Villegas et al., 1999; Hibbett and Thorn, 2001) have shown some agreements between the Jülich (1981), Hawksworth et al. (1995) and Estrada-Torres (1994) classifications.

More precise definitions of certain features, such as spore ornamentation and the shapes of the hilar appendix, have not yet been made. Studies that can focus on these features with detail are of great value in understanding their homology (Patterson, 1982; Roth, 1988; Rieppel and Kearney, 2002). Furthermore, the comparative description of different types of ornamentation, shapes of the hilar appendix and macrochemical reactions could reveal new characters, which may be potentially useful at different taxonomic levels.

Taking into account the various hypotheses regarding the relationships within *Gomphales* drawn partially from spore morphology, this study examined spore structures of representative taxa of *Beenakia*, *Gloeocantharellus*, *Gomphus*, *Kavinia*, *Lentaria*, *Ramaria* and *Ramaricium*. Data on spore morphology available in the literature for this group were also considered. Based on these observations, hypotheses of primary homology are proposed.

Materials and methods

The study looked at around 150 samples of representative taxa of the genera *Beenakia*, *Gloeocantharellus*, *Gomphus*, *Kavinia*, *Lentaria*, *Ramaria* and *Ramaricium*. Thirty of these samples were further selected for microscope scanning, based on the availability of materials and the way of preservation (see list in Table 1). The genera *Delentaria*, *Pseudogomphus* and *Terenodon*, which are monospecific, could not be observed due to the lack of available material. A total of fourteen species, corresponding to ten genera, were

Table 1. List of representative specimens examined in *Gomphaceae sensu lato*.

Taxon	Specimens examined with SEM
Beenakia dacostae	E. Horak 68/448 (L)
Gloeocantharellus purpurascens	F. Tapia 209 (XAL); E. Horak 13977 (TENN)
Gomphus clavatus	G. Guzmán 6845 (ENCB); A.H. Smith s.n. (TENN)
Gomphus floccosus	A. Estrada-Torres 3539 (TLXM); F.R. Bralish 1015 (TENN)
Kavinia alboviridis	A. Estrada-Torres 3058 (TLXM); H.S. Khara 4070 (TENN)
Lentaria surculus	R.E. Halling 5472 (TENN); R.H Petersen 597 (TENN)
Ramaria abietina	R.H. Petersen 2961 (TENN); L. Hernández Díaz 34 (TLXM)
Ramaria cf. botrytis	López & Villegas R.M. 575 (FCME); R. Lamothe & E
	Pérez-Silva 70 (MEXU)
Ramaria cf. fennica	Villegas R.M. 364 (FCME); S. López & Villegas R.M. 437
-	(FCME); Gavito y Santiago 21 (FCME)
Ramaria gracilis	Villegas R.M.1353 (FCME); W.B. Cooke 5000 (TENN)
Ramaria grandis	R.H. Petersen s.n. (TENN); Villegas R.M. 1691 (FCME)
Ramaria rubripermanens	Scates & Petersen 45911 (TENN); A. Estrada-Torres 3244
-	(TLXM); A. Kong Luz 2594 (TLXM)
Ramaria stricta	Smith & R.H.P. 31891 (TENN); L. Guzmán-Dávalos 3595
	(IBUG); E.Pérez, R. Hernández y E. Aguirre (MEXU)
Ramaricium polyporoideum	E.D. Farr & J. Ginns 9732 (NFC); E.D. Farr & J. Ginns 9733
	(NFC)

Herbaria: FCME = Facultad de Ciencias, UNAM, México; TENN = University of Tennessee, Knoxville, EUA; ENCB = Escuela Nacional de Ciencias Biológicas, Instituto Politécnico Nacional; TLXM = Centro de Investigación en Ciencias Biológicas, Universidad Autónoma de Tlaxcala; IBUG = Instituto de Botánica, Universidad de Guadalajara; XAL = Instituto de Ecología, A.C., Xalapa, México; MEXU = Instituto de Biología, UNAM, México; NFC = National Fungus Colection, New York.

investigated (Table 1). These samples are located in the ENCB, FCME, IBUG, L, MEXU, NFC, TENN, TLXM and XAL herbaria.

Techniques proposed by Kotlaba and Pouzar (1964), Marr and Stuntz (1973) and Petersen (1967) were followed for cyanophily observation. Samples placed under a scanning microscope were observed by desiccation of the critical point according to the modified techniques proposed by Espinoza (pers. comm.), using a critical point desiccator BAL-TEC CPD030. The scanning microscope observations (SEM) were made at the Facultad de Ciencias, UNAM, using a JEOL JSM-5310LV electron microscope with an integrated using a magnification range of 5000 to 10000.

Historically, it has not been an easy task to describe spore ornamentation with any accuracy. Therefore, a large number of descriptive terms for spore ornamentation have been introduced, but with little consensus among them. In this study, an attempt was made to employ terminology used in glossaries and dictionaries (Largent *et al.*, 1977; Vellinga, 1988; Hawksworth *et al.*, 1995; Kirk *et al.*, 2001). Images in figures may, however, convey differences

between one type of ornamentation and another with greater clarity. The terminology proposed by Hawksworth *et al.* (1995), Largent *et al.* (1977), Pegler and Young (1971) and Vellinga (1988) was used for the descriptions.

There is no way to ensure that a character possesses phylogenetic information before the similarity patterns between taxa have been analyzed. Therefore, in this study, the characters were evaluated taking into account empirical criteria of similarity, discrete variation between taxa, conjunction, inheritability and independence (De Pinna, 1991; De Luna and Mishler, 1996).

Results

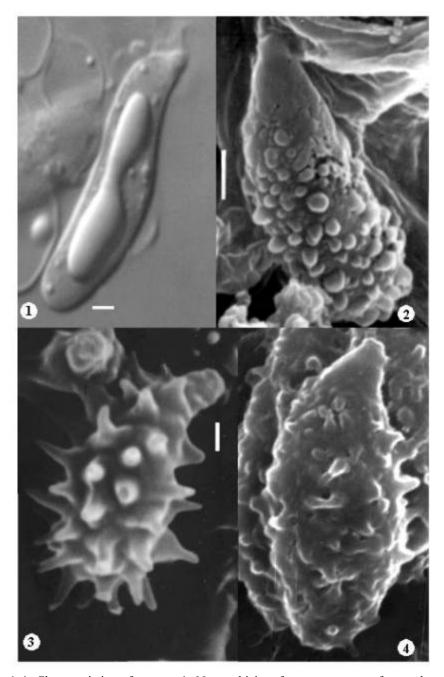
The following are the results obtained in our review of the qualitative characters of spores:

Spore color

Color is one of the characters that, because of its nature, tends to be subjective. Nevertheless, in many basidiomycetes and in our particular case, this feature can be distinguished more clearly in a spore print, which is not always easily obtained. *Gomphales* spores have been described as yellow to ochraceous (Estrada-Torres, 1994), ochraceous-brown, sometimes but rarely straw colored with shades of olive green, cream colored, or colorless (Donk, 1961, 1964; Jülich, 1981). It is evident that color can vary even within the same genus. In *Ramaria*, for example, spores of the subgenus *Echinoramaria* have darker shades than those of *Lentoramaria*.

Under certain circumstances, for example, depending on the features of the sporome and/or environmental conditions, the color of the spore may become either lighter or darker than normal (Largent *et al.*, 1977), rendering color comparison difficult. Since very few spore prints of specimens could be obtained during our study, direct observations were made using a photonic microscope. Despite the use of field guides, descriptions were problematic. Even though the spores examined displayed a gradient of hues, it was evident that the color fluctuated between yellow and brown. This is the case for taxa such as *Beenakia dacostae*, *Gloeocantharellus purpurascens*, *Gomphus clavatus*, *G. floccosus*, *Kavinia alboviridis*, *Ramaria abietina*, *R.* cf. *botrytis*, *R.* cf. *fennica*, *R. gracilis*, *R. grandis*, *R. stricta* and *Ramaricium polyporoideum*.

The only samples to have hyaline spores (observed with a microscope at a magnification of 125×) were *Lentaria surculus*.



Figs. 1-4. Characteristics of spores. **1.** Nomarski interference contrast of smooth spore and hilar appendix obtuse in *Lentaria surculus*. Scanning electron micrograph (SEM) of: **2.** *Ramaria abietina* with spore tuberculate-nodulose and hilar appendix confluent. **3.** Spore echinulate of *Ramaria grandis*. **4.** Spores echinulate and hilar appendix acuminate in *Kavinia alboviridis*. Bars = $1 \mu m$.

Spore shape

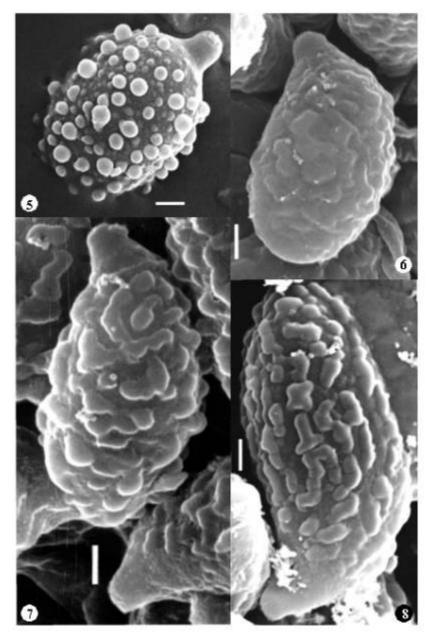
With regard to their symmetry, nearly all taxa observed possessed equilateral spores, with the exception of *Lentaria surculus* (Fig. 1). If the entire spore body is taken into account, the range of spore shapes varied from subglobose to ellipsoid, or more elongated with various gradations in *Ramaria abietina* (Fig. 2), *R.* cf. *botrytis*, *R. grandis* (Fig. 3), *Kavinia alboviridis* (Fig. 4), *Ramaricium polyporoideum* (Fig. 5), *R. gracilis* (Fig. 6), *R. cf. fennica* (Fig. 7), *R. rubripermanens* (Fig. 11), *R. stricta*, *Gomphus clavatus* (Fig. 8), *G. floccosus*, *Gloeocantharellus purpurascens* (Fig. 9), *Beenakia dacostae* (Fig. 10), and to sigmoid and almost cylindrical in *Lentaria surculus* (Fig. 1).

Hilar appendix

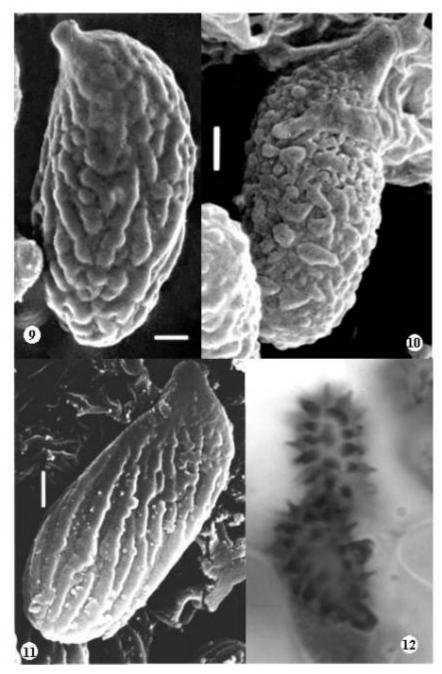
The hilar appendix, which is the protuberance that connects the spore to the sterigma of the basidium, may take a variety of shapes at the base of the spore. According to the terminology proposed by Pegler and Young (1971), these shapes may be: (i) obtuse, in which the base of the spore is broadly round and the hilar appendix is completely different from the rest of the spore; (ii) acuminate, with the base of the spore distinctively attenuated towards the hilar appendix to form a basal prolongation, although the tip is still distinguishable from the rest of the spore, and (iii) confluent, in which the base of the spore gradually becomes thinner. Here, the continuity between the spore and its hilar appendix is more evident and it is very difficult to distinguish between the base of the spore and the hilar appendix. (In this case, the term clavate is also used: elongated spores with a tapering base). With Lentaria surculus and Ramaricium polyporoideum, the hilar appendix was observed to be obtuse (Figs. 1 and 5), whereas Beenakia dacostae, Gloeocantharellus purpurascens, Gomphus clavatus, G. floccosus, Kavinia alboviridis, Ramaria ef. botrytis, R. cf. fennica, R. gracilis, R. rubripermanens and R. stricta had a hilar appendix with an acuminate base (Figs. 4, 6-11). Only Ramaria abietina and R. grandis exhibited a hilar appendix with a confluent base (Figs. 2 and 3).

Spore ornamentation

A great amount of terminology has been introduced to describe spore ornamentation (Largent *et al.*, 1977; Petersen, 1988; Vellinga, 1988; Hawksworth *et al.*, 1995). After making careful comparisons and noting which terms agreed with one another, we arrived at the resulting descriptions:



Figs. 5-8. SEMs of spores. **5.** Ramaricium polyporoideum: tuberculate-nodulose spore and hilar appendix obtuse. **6.** Ramaria gracilis: verrucose ornamentation and hilar appendix acuminate. **7.** Ramaria cf. fennica: hilar appendix acuminate and verrucose spore whith shorter and varied unions. **8.** Gomphus clavatus: verrucose spore. Bars = $1 \mu m$.



Figs. 9-12. SEMs of spores in: 9. Gloeocantharellus purpurascens and 10. Beenakia dacostae with verrucose ornamentation and hilar appendix acuminate. 11. Ramaria rubripermanens: striate spore. 12. Nomarski interference contrast of cyanophilic ornamentation in Ramaria grandis. Bars = 1 μ m.

Smooth

The spore has no ornamentation. Only *Lentaria surculus* (Fig. 1) showed this feature.

Echinulate

The spore surface is covered with broad-based spines, which can be subacute or blunt. The spines may be connected (connate) at their basal portion in the form of reliefs or isthmuses in such a way that these ornamentations are seen as extensions at the spore surface: they do not have an apparent origin at different layers of the wall. This type of ornamentation can be seen in *Kavinia alboviridis* and *Ramaria grandis* (Figs. 4 and 3).

Tuberculate-nodulose

The spore surface has prominent excrescences, such as individual warts, more or less equidistant from, or rarely connected to one another. This type of ornamentation was observed only in *Ramaria abietina* and *Ramaricium polyporoideum* (Figs. 2 and 5).

Verrucose

The spore surface is covered by obvious warts or round lobes. These ornamentations can be connected to each other in different degrees. In taxa such as *Gloeocantharellus purpurascens* (Fig. 9) and *Gomphus floccosus*, the connection is quite extensive, forming waves or irregular 'cords', although not forming striae. In *Beenakia dacostae*, *Gomphus clavatus*, *Ramaria* cf. *fennica*, *R. gracilis* and *R. stricta*, on the other hand, the warts tend to have shorter and varied connections (Figs. 10, 8, 7 and 6). They can also be isolated. Species such as *R. gracilis* and *R. stricta* exhibit smaller and more separate warts in comparison to the other taxa considered in this group.

Striate

The spore ornamentation is organized in shallow lines or grooves, which apparently arise from the external layer(s) of the wall, following a divergent spiral pattern that runs from the abaxial to the adaxial part of the spore. A few round warts can be seen in the frontal portion, especially in immature spores. This suggests that the ornamentation might be formed from the union of these warts or, more likely, we are dealing with two different ornamental elements. This is a characteristic of *Ramaria* cf. *botrytis* and *R. rubripermanens* (Fig. 11). When considering the fundamental principles for the selection of primary homologies, the collective presence of two discrete states of a character in the same organism (isolated warts and continuous striate) allows us to infer that

there are two independent characters instead of just one (De Pinna, 1991; De Luna and Mishler, 1996; Hawkins *et al.*, 1997). It is considered, therefore, that the striate condition is independent from the verrucose one.

In *Kavinia alboviridis*, for example, even though the spore ornamentation is mainly characterized by sharp projections, it also tends to present round tips, similar to warts. In *Ramaria abietina*, some nodules or tubercules tend to become slightly slender towards the tip. It is evident that these elements involve variations at another level. For this reason, these patterns were considered as more evident modalities at a generic level.

Cyanophilic reaction

According to Kotlaba and Pouzar (1964), Petersen (1967) and Largent *et al.* (1977), a spore is cyanophilic when its wall absorbs a great concentration of cotton blue, acquiring a more intense color than the cytoplasm. When the wall or the ornamentation stands in distinct contrast to the internal portion of the spore, the former are called cyanophilic.

With the exception of *Lentaria surculus*, all the remaining taxa studied in this work are cyanophilic. Nevertheless, some differences do exist in the site of reaction. Spores of *Beenakia dacostae*, *Gloeocantharellus purpurascens*, *Gomphus clavatus*, *G. floccosus*, *Kavinia alboviridis*, *Ramaria abietina*, *R.* cf. *botrytis*, *R.* cf. *fennica*, *R. gracilis*, *R. grandis*, *R. rubripermanens*, *R. stricta* and *Ramaricium polyporoideum* absorb the dye rapidly but the reaction takes place only in the ornamentation; the wall does not absorb the dye (Fig. 12). The extent of the cyanophilic reaction varies from species to species depending on the ornamentations dimension and density. The reaction is very evident in species such as *Beenakia dacostae*, *Gomphus clavatus*, *G. floccosus*, *Kavinia alboviridis*, *Ramaria abietina*, *R.* cf. *fennica* and *R. grandis*, among others, and less evident in *R. gracilis* and *R. stricta*. In *Ramaria* cf. *botrytis* and *R. rubripermanens*, the cyanophilic reaction follows the pattern of the 'grooves' more than the crests.

In general, it can be said that the taxa showing a conspicuous ornamentation under the light microscope clearly reacted strongly to cotton blue, whereas those with smooth spores or inconspicuous ornamentation had a less noticeable reaction.

Discussion

Spore characters are widely used in classification across a range of basidonmycetes (e.g. Desjardin *et al.*, 2003; Das *et al.*, 2004). In this study we

examined the stability of spore characters in the *Glomerales*, but the results may apply across a broad range of fungi.

Spore color

The results obtained in this study show that, although color was described in a qualitative manner, it could vary considerably according to the species and their developmental stages. Methodological problems have not allowed us to consider this variation in descriptive or quantifiable terms.

Spore shape

If we consider spore shape to be a qualitative feature, it is difficult to represent the great variation in general shapes as discrete character states. The transformation of spore shapes into quantitative characters in morphometric studies may enable us to use them at the desired level explained above. Nevertheless, using characters such as symmetry and the shape of the spore base, it is interesting to note that, according to Domínguez de Toledo (1994) and Largent *et al.* (1977), it is possible to identify taxa such as *Lentaria surculus*, which present a clearly un-equilateral symmetry, while others consistently exhibit a more regular (equilateral) symmetry.

Hilar appendix

It is evident that the hilar appendix has inherent variations in its shape; however, the only available data obtained in this study refers to external morphology and position in relation to the rest of the spore. In general, we know that the hilar appendix is always present at the base of the basidiospores. But, unlike with spore ornamentation, it is not known with certainty if this structure is made up of only one or several wall layers. The consensus at the moment is that it is probably made up of the basic structure of spore layers, since a thinner wall is generally observed (Keller, 1992, 1997). Despite this, at present, it is difficult to distinguish if a hilar appendix with an obtuse base is ultrastucturally and ontogenetically different from one that is acuminate or confluent. There is even the possibility that this feature is independent from the structure. The current evidence allows us to acknowledge only that there is a positional similarity with different base shapes. The base shape of the spore and the hilar appendix seen as a hypothesis of 'primary homology' (De Pinna, 1991) might suggest that they vary altogether (round base with obtuse hilar appendix and a pointed base with an acuminate hilar appendix). Nevertheless,

the conjunction of the spore base, the sharp tip and a confluent hilar appendix in *Ramaria abietina* and *R. grandis* leads us to suggest that they are non-homologous features, implying at least two events of change or, in other words, two independent characters.

Spore ornamentation

It is evident that ornamentation exhibits great variation. Despite the existence of different patterns, these may be present without distinction in some genera such as *Ramaria*. Indeed, it is within the *Gomphales* that the greatest diversity of spore ornamentation can be seen: from the verrucose to echinulate.

Petersen (1981) classified spore ornamentation in *Ramaria* subgenus *Echinoramaria* into five different types: (i) with long and sharp spines, (ii) a mixture of long, sharp spines and shorter elements, (iii) with telephoroid ornamentation, (iv) with truncate ('volcanic') spines, and (v) with rounded warts. If this classification is examined in detail, comparing the images of our results with those presented by Petersen (1981), it becomes evident that types 1 to 4 correspond to what we consider as tall ornamentations with projections at the tip, but with variations in dimension or shapes of the tips (volcanic or telephoroid).

On the other hand, it is evident that, in the case of *Ramaria abietina* (Fig. 2), as is generally assumed in the descriptions arrived at using a light microscope, the ornamentation is in the form of round warts, not sharp spines, a feature which Petersen (1981) also revealed in *Ramaria myceliosa*. Both of these *Ramaria* are classified in the *Flaccidae* section of the subgenus *Echinoramaria*. Petersen (1981) also speculated on the possibility that the sections *Flaccidae* and *Dendrocladium* have very little in common, considering, among other aspects, spore ornamentation. Although the cladistic hypotheses currently proposed for the *Gomphales* (Villegas *et al.*, 1999; Humpert *et al.*, 2001) suggest *Ramaria* to be polyphyletic, there has been no additional study to clarify the generic classification in this group. It is important to point out that Villegas *et al.* (1999) showed that the two species of *Echinoramaria* (*R. abietina* of section *Flaccidae* and *R. grandis* of section *Dendrocladium*) are not grouped using the criteria presented in this study.

A strong resemblance in spore ornamentation has been observed for various species of *Gomphus* and *Ramaria* (Corner, 1950; Eriksson, 1954; Petersen, 1971a). It is important to point out that this similarity in spore ornamentation is indeed surprising. Up to now, it has not been suggested that this similarity corresponds to any discrete pattern or definitive grouping, but

has been observed without distinction in several species traditionally classified in different sub-generic levels. Examples include *G. clavatus* and *R.* cf. *fennica* (Petersen, 1971a, 1981, 1988; Pegler and Young, 1981), *Gomphus guadelupensis* and *G. retisporus* and species of subgenus *Echinoramaria* such as *R. cristatospora* and *R. pancaribbea*, among others. We can also see similarities between species such as *Gloeocantharellus purpurascens* and *Gomphus floccosus*, or between *Gomphus clavatus* and *Ramaria flava*, *R. flavobrunescens*, and *R. fumigata* (Corner, 1966; Petersen, 1967, 1971a; Keller, 1997). For these latter works, hypotheses of intergeneric relationships have been based on these features.

Ornamentation has been considered equally important in the differentiation of the subgenera within *Ramaria* (Corner, 1970; Marr and Stuntz, 1973; Petersen, 1988). Researchers using a light microscope described the spores of the subgenus *Lentoramaria* as having a 'lighter' ornamentation than those of subgenus *Laeticolora* (Petersen, 1975). The results of this investigation show that there is no clear distinction that consistently separates these subgenera because the form and position of the ornamentation are very similar. Moreover, we must bear in mind that the spore dimensions in the subgenus *Lentoramaria* are smaller than those of the subgenus *Laeticolora*. In addition, this group has less conspicuous ornamentation, as pointed out by Keller (1997) in his work with *Ramaria flavosalmonicolor*. For the subgenus *Ramaria*, on the other hand, the spores have a homogeneous ornamentation pattern, with a conspicuous longitudinal arrangement of parallel striae, totally or slightly discontinuous, with occasional isolated warts.

Corner (1950) also suggested a possible relationship between *Lentaria* and some taxa of the *Ramaria gracilis* group, considering, among other aspects, the scarce spore ornamentation in the latter as an indication. As is shown in Figs. 1 and 6, combined with descriptions by Petersen (1967, 1968, 1971b, 1975), for instance, the spores in *Lentaria* are smooth, whereas the species of the group of *Ramaria gracilis* present ornamented spores.

The same ornamentation pattern found in our investigation for *Beenakia dacostae* is also found in the study of Parmasto and Ryvarden (1990) on *Beenakia fuliginosa* and *B. informis*. Therefore, it is likely that is a group with spores of homogeneous ornamentation.

In recent decades, spore wall construction has received a great deal of attention (Keller, 1997). Studies carried out on this and other groups of hymenomycetes have shown that spores may have a variable number of layers in the walls (between three and five) and that the ornamentation may originate from different strata (Clémençon, 1997; Keller, 1997). The results obtained by Jülich and Star (1983) show that the spore wall in *Beenakia dacostae* is

identical to the structure known for the species of *Gomphus* and *Ramaria* studied by Perreau (1967), where it became evident that the ornamentation arises from the outermost layer of the wall, whose width is greater than the inner layers. Similar characteristics are shown by Vázquez *et al.* (1995) for some species of *Ramaria*.

Authors such as Perreau (1967), Clémençon (1970, 1973) and Keller (1986, 1997) have demonstrated that the layers of the spore wall are not always arranged in the same order. Keller (1997), studying some species of *Gomphus* and *Ramaria*, described the spore wall in *Gomphaceae* (sensu Donk) as thick and irregular. It was constituted of (i) a coriotunica, which is the fundamental basis of all spores, which could be granular, fibrilar or stratified; (ii) a tunica, represented by opaque material; (iii) a podostratum, seen as a transparent thin layer which does not react to iodine; and (iv) a mucostratum, which reacts to cotton blue.

In Gomphaceae, it has been suggested that the ornamentation arises from the mucostratum layer (Keller, 1986). In absence of ontogenetic studies that would allow us to recognize the order of formation and aggregation of the layers of the spore wall, and in view of the fact that up to this point this process is known in only relatively few taxa of Gomphales, there is not enough evidence to propose homologies between these characters or to establish correlations between the ornamentation patterns. More studies of this group, using electron micrographs, may enable us to understand more about the similarity observed in these ornamentation patterns.

All the data obtained for this study lead us to conjecture that, at least within *Gomphales*, there are four hypotheses of homology: (i) spores without ornamentation; (ii) ornamentation with pointed projections that can be differentiated into conical shapes with or without anastomosed bases; (iii) round ornamentation with the following distinguished states: (a) tuberculate-nodulose and (b) verrucose; and (iv) striate ornamentation. At a first glance, this last character could be considered as belonging to 2. However, the presence of striae and warts in the same spore lead us to believe that they are two different characters. It is clear, therefore, that it is not feasible to consider the whole ornamental character under only one hypothesis of transformational homology.

Cyanophilic reaction

The cyanophilic reaction of the spore ornamentation was considered as a primary taxonomic feature by Donk (1964). From this perspective, authors such as Petersen (1967) considered that *Gomphus* and some species of

Ramaria are closely related. A similar connection has been suggested for *Beenakia* and *Ramaria* (Maas Geesteranus, 1963).

Petersen (1967) observed that the spore surface in *Ramaria grandis* is conspicuously more colored than the areas between the spines. Marr and Stuntz (1973), in a comparative study of spores of *Ramaria* treated with cotton blue, noted that the ornamentation was always cyanophilic, but that it may or may not be distinct from the rest of the wall. In *Ramaria* cf. *botrytis*, the cyanophilic reaction is less conspicuous. Petersen and Pearman (1973) showed that, contrary to what happens in species of other *Ramaria* subgenera, the cyanophilic reaction is lighter in the crests of the striae and more intense in the areas between those striae, a feature that, from our perspective, is probably related to the number and the thickness of the wall layers.

It is clear that, within *Gomphales*, there is a group of taxa whose spores react with cotton blue, albeit not in a uniform manner (Marr and Stuntz, 1973; Petersen and Pearman, 1973). But, as with ornamentation, we are not convinced that the variations in the cyanophilic reaction are due to different ultrastructural, chemical or ontogenetic causes. Because of this, the deductive, inferential process allows us to consider only the presence or absence of cyanophyly as a likely taxonomical character at this level. In preliminary studies (Villegas et al., 1999), cyanophily has been postulated as a synapomorphy for the group including Beenakia, Gloeocantharellus, Gomphus, Kavinia, Ramaria and Ramaricium. Here the polarization of characters, by means of external groups, suggests a phylogenetic transformation from absence of cyanophily towards a condition of derived possibly phylogenetically correlated with ornamentation and spore color. These results suggest that cyanophily could be interpreted as a synapomorphy that could unite the aforementioned genera into a monophyletic group.

Finally, it is clear that all these hypotheses must be evaluated by their phylogenetic congruence with other characters, as proposed by De Pinna (1991), Roth (1988) and De Luna and Mishler (1996), includign mating studies where feasible (e.g. Peterson and Hughes, 2003; Mata *et al.*, 2004) and molecular phylogenetics (e.g. Keirle *et al.*, 2004; Zhang *et al.*, 2004).

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References

- Brower, A.V.Z. (2000). Homology and the inference of systematic relationships: some historical and philosophical perspectives. In: *Homology and Systematics* (eds. R. Scotland and R.T. Pennington). Taylor & Francis Inc, London: 10-21.
- Bryant, H.N. (1989). An evaluation of cladistic and character analysis as hypothetico-deductive procedures, and the consequences for character weighting. Systematic Zoology 38: 214-227
- Clémençon, H. (1970). Bau der Wände der Basidiosporen und ein Vorschlag zur Benennung ihrer Schichten. Zeitschrift fur Pilzkunde 36: 113-133.
- Clémençon, H. (1973). Die Wandstrukturen der Basidiosporen. III *Cortinarius* und *Dermocybe*. Zeitschrift für Pilzkunde 39: 121-144.
- Clémençon, H. (1997). Anatomie der Hymenomyceten. Benteli Druck, Schweiz.
- Corner, E.J.H. (1950). A monograph of *Clavaria* and allied genera. Annals of Botany Memmory 1: 1-740.
- Corner, E.J.H. (1966). Cantharelloid Fungi. Oxford University Press, Oxford.
- Corner, E.J.H. (1970). Supplement to a monograph of Clavaria an allied genera. Cramer, Lehre.
- Das, K., Sharma, J.R. and Montoya, L. (2004). *Lactarius (Russulaceae)* in Kumoan Himalaya. !. New species of subgenus *Russularia*. Fungal Diversity 16: 23-33.
- De Luna, E. and Mishler, B.D. (1996). El concepto de homología filogenética y la selección de caracteres taxonómicos. Boletín de la Sociedad Botánica de México 59: 131-146.
- De Pinna, M.C.C. (1991). Concepts and tests of homology in the cladistic paradigm. Cladistics 7: 367-394.
- Desjardin, D.E., Boonpratuang, T. and Hywel-Jones, N. (2003). New spinose species of *Mycena* in sections Basipedes and Polyadelphia from Thailand. Fungal Diversity 12: 7-17.
- Domínguez de Toledo, L.S. (1994). Suggestions for describing and illustrating fungal spores. Mycotaxon 52: 259-270.
- Donk, M.A. (1961). Four New Families of Hymenomycetes. Persoonia 1: 405-407.
- Donk, M.A. (1964). A Conspectus of the Families of Aphyllophorales. Persoonia 3: 199-324.
- Eriksson, J. (1954). *Ramaricium* n. gen., a corticioid member of the *Ramaria* group. Svensk Botanisk Tidskriff 48: 188-198.
- Estrada-Torres, A. (1994). La Familia Gomphaceae (Aphyllophorales: Fungi) en el estado de Tlaxcala. Tesis Doctoral. Escuela Nacional de Ciencias Biológicas del IPN, México.
- Hall, B.K. (1994). *Homology. The Hierarchical Basis of Comparative Biology*. Academic Press, London.
- Hawkins, J.A. (2000). A survey of primary homology assessment: different botanists perceive and define characters in different ways. In: *Homology and Systematics* (eds. R. Scotland and R.T. Pennington). Taylor & Francis Inc, London: 22-53.
- Hawkins, J.A., Hughes, C.E. and Scotland, R.W. (1997). Primary homology assessment, characters and character states. Cladistics 13: 275-283.
- Hawksworth, D.L., Kirk, P.M., Sutton, B.C. and Pegler, D.N. (1995). *Ainsworth & Bisby's Dictionary of the Fungi*. 8th ed. International Mycological Institute, CAB International, Wallingford.
- Heim, R. (1970). Breves diagnoses latinae novitatum genericarum specificarumque nuper descriptarum. Revue de Mycologie 34: 343-347.

- Hibbett, D., Pine, E.M., Langer, E., Langer, G. and Donoghue, M.J. (1997). Evolution of gilled mushrooms and puffballs inferred from ribosomal DNA sequences. Proceedings of the National Academy Sciences 94: 12002-12006.
- Hibbett, D.S. and Thorn, R.G. (2001). Basidiomycota:Homobasidiomycetes. In: *The Mycota*, *vol. VII part B, Systematics and Evolution* (eds. D.J. McLaughlin and P.A. Lemke). Springer Verlag, Berlin: 121-168.
- Humpert, A.J., Muench, A.J., Giachini, M.A., Castellano, M.A. and Spatafora, J.W. (2001). Molecular phylogenetics of *Ramaria* and related genera: evidence from nuclear large subunit and mitochondrial small subunit rDNA sequences. Mycologia 93: 465-477.
- Jülich, W. (1981). Higher taxa of Basidiomycetes. J. Cramer, Vaduz.
- Jülich, W. and Star, W. (1983). Ultrastructure of basidiospores I. Beenakia. Persoonia 12: 67-74.
- Keller, J. (1986). Ultrastructure des parois sporiques des Aphyllophorales. Mycolgia Helvetica 2: 1-34.
- Keller, J. (1992). Ultraestructure de la paroi sporique des Heterobasidiomycetes I. Persoonia 14: 377-387.
- Keller, J. (1997). *Atlas des Basidiomycetes*. Union Des Societes Souisses De Mycologie, NeuchCtel.
- Keirle, M.R., Hemmes, D.E. and Desjardin, D.E. (2004). Agaricales of the Hawaiian Islands. 8. Agaricaceae: *Coprinus* and *Podaxis*; Psathyrellaceae: *Coprinopsis*, *Coprinellus* and *Parasola*. Fungal Diversity 15: 33-124.
- Kirk, P.M., Cannon, P.F., David, J.C. and Stalpers, J.A. (2001). *Ainsworth & Bisby's Dictionary of the Fungi*. International Mycological Institute, CAB International, Wallingford.
- Kotlaba, F. and Pouzar, Z. (1964). Preliminary results on the staining of spores and other structures of Homobasidiomycetes in cotton blue and its importance for taxonomy. Feddes Repert 69: 131-142.
- Kühner, R. (1976). La notion d'espece chez les champignons superieurs. Bulletin de la Societe Linneenne de Lyon 45: 19-48.
- Largent, L., Johnson, D. and Watling, R. (1977). How to Identify Mushrooms to Genus III.

 Microscopic features. Mad River Press, Eureka.
- Maas Geesteranus, R.A. (1963). Hyphal structures in Hydnums. III. Proceedings of the Koninklijke Nederlandse Akademie van Wetenschappen Series C 66: 437-446.
- Maas Geesteranus, R.A. (1971). Hydnaceous Fungi of the eastern Old World. Verh. K. Ned. Akad. Wet. Afd. Natuurk., Tweede Reeks 60: Amsterdam.
- Mabee, P.M. (1989). Assumptions underlaying the use of ontogenetic sequences for determining character state order. Transactions of the American Fisheries Society 118: 151-158.
- Marr, C.D. and Stuntz, D.E. (1973). Ramaria in western Washington. J. Cramer, Lehre.
- Mata, J.L., Halling, R.E. and Petersen, R.H. (2004). New species and mating system reports in *Gymnopus (Agaricales)* from Costa Rica. Fungal Diversity 16: 113-129.
- Parmasto, E. and Ryvarden, L. (1990). The genus *Beenakia* (Gomphaceae, Aphyllophorales). Windalhia 18: 35-42.
- Patterson, C. (1982). Morphological characters and homology. In: *Problems of Phylogenetic Reconstruction* (eds. K.A. Joysey and A.E. Friday). Academic Press, London: 21-74.
- Pegler, D.N. and Young, T.W.K. (1971). *Basidiospora morphology in the Agaricales*. J. Cramer, Lehre.
- Pegler, D.N. and Young, T.W.K. (1979). The gasteroid Russulales. Transactions of the British Mycological Society 72: 353-388.

- Pegler, D.N. and Young, T.W.K. (1981). A natural arragement of the Boletales, with reference to spore morphology. Transactions of the British Mycological Society 76: 103-146.
- Perreau, B.J. (1967). Recherches sur la differenciation et la structure de la paroi sporale chez les Homobasidiomycetes á spores ornèes. Annales of Sciences Naturals of Paris 12: 639-746.
- Petersen, R.H. (1967). Evidence on the interrelationships of the families of clavarioid fungi. Transactions of the British Mycological Society 50: 641-648.
- Petersen, R.H. (1968). Notes on cantharelloid fungi I. *Gomphus* S.F. Gray and some clues to the origin of ramarioid fungi. Journal of Elisha Mitchell Scientific Societty 84: 373-381.
- Petersen, R.H. (1971a). *The genera Gomphus and Gloeocantharellus in North America*. J. Cramer, Lehre.
- Petersen, R.H. (1971b). Notes on Clavarioid Fungi-IX. Addendum to *Clavulinopsis* in North America. Persoonia 6: 219-229.
- Petersen, R.H. (1975). *Ramaria* subgenus *Lentoramaria* with emphasis on North American Taxa. Bibliotheca Mycologica 43: 1-161.
- Petersen, R.H. (1981). Ramaria subgenus Echinoramaria. Bibliotheca Mycologica 79: 1-261.
- Petersen, R.H. (1988). *The Clavarioid Fungi of New Zealand*. Science Information Publishing Centre, New Zealand.
- Petersen, R.H. and Pearman, W.R. (1973). Spore ornamentation in *Ramaria* as depicted by scanning electron micrographs. Persoonia 7: 289-292.
- Petersen, R.H. and Hughes, K.W. (2003). Phylogenetic examples of Asian biodiversity in mushrooms and their relatives. Fungal Diversity 13: 95-109.
- Pine, E.M., Hibbett, D.S and Donoghue, M.J. (1999). Phylogenetic relationships of *cantharelloid* and *clavarioid* Homobasidiomycetes based on mitochondrial and nuclear DNA sequences. Mycologia 91: 944-963.
- Rieppel, O. and Kearney, M. (2002). Similarity. Biological Journal of the Linnean Society 75: 59-82.
- Roth, V.L. (1988). The biological basis of homology. In: *Ontogeny and systematics* (eds. C.J. Humpriers). Columbia University Press, New York: 1-26.
- Vázquez, G., Cano-Canchola, C., Sosa, L., Obregón, A., Flores-Carreón, A., Herrera-Estrella, A., Villegas, M. and Cifuentes J. (1995). Classical and molecular studies on edible species of *Ramaria* from México. In: *Science and Cultivation of Edible Fungi* (eds. T.J. Elliot). A.A. Balkema, Rotterdam: 147-152.
- Vellinga, E.C. (1988). Glosary. In: *Flora Agaricina Neerlandica. Vol. 1* (eds. C. Bas, T.H.W. Kuyper, M.E. Noordeloos and E.C. Vellinga). A.A. Balkema, Roterdam: 54-64.
- Villegas, M., De Luna, E., Cifuentes, J. and Estrada-Torres, A. (1999). Phylogenetic studies in Gomphaceae *sensu lato* (Basidiomycetes). Mycotaxon 70: 127-147.
- Zhang, L.F., Yang, J.B., Yang, Z.L. (2004). Molecular phylogeny of eastern Asian species of *Amanita* (Agaricales, Basidiomycota): taxonomic and biogeographic implications. Fungal Diversity 17: 219-238.

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